NOTE

Syntheses of ¹⁸F-Labeled Reduced Haloperidol and ¹¹C-Labeled Reduced 3-N-Methylspiperone

Hayden T. Ravert, Robert F. Dannals, Alan A. Wilson, Dean F. Wong, and Henry N. Wagner, Jr.

Divisions of Nuclear Medicine and Radiation Health Sciences The Johns Hopkins Medical Institutions Baltimore, Maryland 21205-2179

SUMMARY

¹⁸F-Labeled reduced haloperidol and ¹¹C-labeled reduced 3-Nmethylspiperone were synthesized in a convenient and quantitative one step reduction from ¹⁸F-labeled haloperidol and ¹¹C-labeled Nmethylspiperone, respectively. Both products were purified by semipreparative HPLC and were obtained at high specific activity and radiochemical purity.

Key Words: Reduced haloperidol, reduced methyl-spiperone, carbon-11, fluorine-18, positron emission tomography

INTRODUCTION

Since 1978 the butyrophenone neuroleptic haloperidol has been known to undergo metabolic reduction of its ketone to form the corresponding chiral secondary alcohol, reduced haloperidol (1). *In vitro* studies with rat and guinea pig liver microsomes have shown an interconversion of haloperidol and reduced haloperidol (2). This interconversion has also been observed in normal volunteers (3). Reduced haloperidol is known to cross the blood-brain barrier. Although reduced haloperidol has some uptake in the striatum, it is relatively ineffective as an antagonist of central dopamine receptors in the rat (4). A study of schizophrenics who did not respond to haloperidol therapy indicated high reduced haloperidol levels (5,6). Recently, a reduced metabolite of another butyrophenone, 3-N-methylspiperone, has been observed in guinea pigs (7). Positron emitting butyrophenone neuroleptic radiotracers for studying dopamine D-2 receptors would be expected to exhibit the same metabolic reduction as their stable counterparts. The radiosynthesis of the reduced form of the neuroleptic may help to understand the metabolic fate of the neuroleptic radiotracer and aid in mathematical modeling of receptor concentrations. This paper reports the facile reduction of ¹⁸F-haloperidol to reduced ¹⁸F-haloperidol and 3-N-[¹¹C-methyl]spiperone to reduced 3-N-[¹¹C-methyl]spiperone.

RESULTS

¹⁸F-Haloperidol was synthesized according to the method of Shiue *et al* (8,9) with minor modifications that are described in the experimental section. Crude ¹⁸F-haloperidol was reduced by excess NaBH₄ in methanol. Heating at 80° C for one minute resulted in quantitative conversion to reduced ¹⁸F-haloperidol. Semipreparative HPLC using a silica stationary phase and a reverse mobile phase provided an suitable separation of ¹⁸F-haloperidol (k' = 5.8) and reduced ¹⁸F-haloperidol (k' = 7.1). A minor mass peak (k' = 6.4), possibly due to reduced nitrohaloperidol, lowered the apparent specific activity (10) to between 100 - 1500 mCi/µmole. The synthesis was completed in 88 minutes with an average yield of 10% (decay corrected from starting fluoride).

¹¹C-Labeled reduced 3-N-methylspiperone was synthesized by two slightly different methods. In method 1, 3-N-[¹¹C-methyl]spiperone (k' = 6.8) was purified by semipreparative reverse phase HPLC. The radiolabeled product was well separated from the unreacted spiperone (k' = 3.5) providing specific activities above 2000 mCi/µmole. The 3-N-[¹¹C-methyl]spiperone was dissolved in saline and NaBH₄ added. After heating at 80° C for two minutes, a quantitative conversion to reduced 3-N-[¹¹C-methyl]spiperone was obtained. The conversion to the reduced species was slower than observed with the reduction of ¹⁸F-haloperidol; this is possibly due to the larger volume of solvent used in the radiocarbon synthesis. After dilution and filtration, the solution was determined to be chemically and radiochemically pure by analytical HPLC with specific activities still above 2000 mCi/µmole. The synthesis was completed in 20 minutes with a yield of 33% (decay corrected from ¹¹CH₃I).

Method 2 involved the reduction of crude 3-N-[¹¹C-methyl]spiperone in its original DMF solution using excess NaBH₄ and methanol. After heating at 80° C for one minute, the reduction was complete. The same semipreparative HPLC conditions as used in method 1 rendered a poor separation of reduced 3-N-[¹¹C-methyl]spiperone (k' = 2.7) and another mass peak presumably reduced spiperone (k' = 1.8). The apparent specific activity for method 2 was an order of magnitude lower compared with the product specific activity from method 1. The synthesis was completed in 19 minutes with a yield of 11% (decay corrected from ¹¹CH₃I).

EXPERIMENTAL

Spiperone and N-methylspiperone were synthesized according to the references cited and were analytically pure (11,12). Reduced haloperidol was obtained from Research Biochemicals Inc., Natick, MA. Elemental analyses were performed by Atlantic Microlabs Inc., Norcross, GA. NMR and IR spectra were obtained on an IBM NR80 and a Perkin Elmer 399B instrument respectively. High performance liquid chromatographic analysis and purification were performed with two Waters 590EF HPLC pump, an in-line fixed wavelength (254 nm) uv detector, and a single two inch NaI crystal radioactive detector. HPLC chromatograms were recorded using two Hewlett-Packard 3390A integrators. HPLC semipreparative purifications were performed on either: column A - Alltech Econosil 10 μ C-18 column (1.0 x 25 cm) or Column B - Beckmann 10 μ silica column (1.0 x 25 cm). A dose calibrator (Capintec 12R) was used for all radioactivity measurements.

Reduced ¹⁸**F**-Haloperidol. The first portion of the preparation is a modification of the reported synthesis of ¹⁸F-haloperidol (8,9). The modifications include the use of K₂CO₃/Kryptofix instead of Cs₂CO₃ and changes in the solvents used in the alkylation step. No carrier added ¹⁸F-fluoride, prepared by proton bombardment of ¹⁸O-H₂O (95 - 99%) using a Scanditronix MC-16F biomedical cyclotron, was concentrated on a short column of anion exchange resin (Dowex 1-X8, 200-400 mesh, hydroxide form). ¹⁸F-Fluoride was eluted from the column using K₂CO₃ (2.3 mg, 17 µmoles) dissolved in 300 µL water into a conical glass vial containing Kryptofix 2.2.2 (13 mg, 34 µmoles). The fluoride was dried by repeated azeotropic distillation with CH₃CN at 120° C under argon gas flow.

After heating for an additional 3 minutes, cyclopropyl p-nitrophenyl ketone (2 mg, 10 μ moles) dissolved in 200 μ L of dry freshly distilled DMSO was added and the vial was heated at 120° C for 10 minutes. After cooling to room temperature, 2 mL of concentrated HCl:methanol (1:1) was added to the vial. The vial was heated at 120° C for 3 minutes, then it was cooled and 3 mL of H₂O added. The resulting solution was applied to a C-18 SEP-PAK and the SEP-PAK was washed with 4 mL of H₂O and 0.5 mL of pentane.

The intermediate product was eluted from the SEP-PAK with an additional 5 mL of pentane through a K_2CO_3 column into a conical vial containing 4-(4-chlorophenyl)-4-hydroxypiperidine (3 mg, 14 µmoles) and KI (8 mg, 48 µmoles) dissolved in 200 µL of DMF. Volatile solvents were removed under an argon flow and the resulting DMF solution was heated at 120° C for 10 minutes. After cooling, 500 µL of methanol and 5 mL of H₂O were added. The mixture was passed through a C-18 SEP-PAK which was rinsed with 4 mL H₂O. The crude product was eluted from the SEP-PAK with 5 mL methylene chloride and passed through a K₂CO₃ column into a conical vial, and evaporated to dryness under a stream of argon.

Solid NaBH₄ (1 mg, 26 μ moles) and 200 μ L of methanol were added. The vial was sealed and heated to 80° C for 2 minutes. HPLC buffer solution (400 μ L) was added and the mixture was injected onto semi-preparative column B (70% CH₃CN / 30% 0.004 M NH₄H₂PO₄, flow rate of 11 mL/min). The reduced ¹⁸F-haloperidol (retention time = 7.7 minutes, k' = 7.1) was collected, evaporated to dryness under reduced pressure, redissolved in 7 mL of sterile saline, and passed through a sterile 0.2 μ filter into a sterile evacuated vial. Sterile 8.4% sodium bicarbonate (3 mL) was added to the vial. The determination of chemical and radiochemical purity was performed using a Waters Novapak 4 μ C-18 column (0.39 x 15 cm) and mobile phase of 30% CH₃CN / 70% 0.1 M NH₄HCO₂ at flow rate of 2 mL/min. Under these conditions, reduced ¹⁸F-reduced haloperidol and haloperidol have k's of 2.5 and 5.2, respectively.

Reduced 3-N- $[^{11}C$ -methyl]spiperone. Method 1. The synthesis of this tracer involved the production of 3-N- $[^{11}C$ -methyl]spiperone according to the published procedure (13). Briefly, $^{11}CH_3I$ produced from $^{11}CO_2$ was bubbled

through a DMF (200 μ L) solution of spiperone (1 mg, 25 μ mole). Tetrabutylammonium hydroxide (5 μ L, 0.4 M in H₂O) was added to the vial and the vial was heated at 80° C for 1 minute. The intermediate product, 3-N-[¹¹C-methyl]spiperone (retention time = 7.0 minutes, k' = 6.8), was purified by semipreparative HPLC (column B, 50% CH₃CN / 50% 0.1 M NH₄HCO₂, flow rate of 11 mL/min) and reconstituted in 1 mL of sterile saline.

Solid NaBH₄ (2 mg, 52 μ mole) was added and the flask was heated to 80° C for 2 minutes. The solution was diluted with 6 mL of sterile saline and passed through a sterile 0.2 μ filter into a sterile evacuated vial. Sterile 8.4% sodium bicarbonate (3 mL) was added to the vial. An Alltech Econosil 10 μ C-18 analytical (0.46 x 25 cm) column (50% CH₃CN/50% 0.1 M NH₄HCO₂) at a flow rate of 4 mL/min) was used for radiochemical and chemical purity determinations. Under these conditions, ¹¹C-reduced N-methylspiperone and N-methylspiperone have k's of 0.8 and 1.5, respectively.

Reduced 3-N-[¹¹C-methyl]spiperone. Method 2. $3-N-[^{11}C-Methyl]$ spiperone was synthesized in DMF as described in method 1. To the crude mixture was added NaBH₄ (1 mg, 26 µmole) and methanol (200µL). After heating the solution at 80° C for 1 minute, the mixture was diluted with HPLC mobile phase (200 µL) and applied to column B (50% CH₃CN / 50% 0.1 M NH₄HCO₂, flow rate: 11 mL/min). The product (k' = 2.7) was collected, evaporated to dryness under reduced pressure, and reconstituted in 7 mL of sterile saline. Following Millipore filtration into a sterile evacuated vial, sterile 8.4% sodium bicarbonate (3 mL) was added. The analytical HPLC conditions were the same as in method 1 above.

Reduced N-methylspiperone - (R,S)-8-[4-(4-fluorophenyl)-4hydroxybutyl]-1-phenyl-1,3,8-triazaspiro[4.5]decan-4-one. Sodium borohydride (20 mg, 530 µmole) was added to 3-N-methyl-spiperone (100 mg, 240 µmole) dissolved in 4 mL of methanol. After 5 minutes stirring, water (5 mL) was added and a white precipitate formed. The mixture was extracted with methylene chloride (3 x 5 mL). The organic layer was washed with water (5 mL), dried over Na₂SO₄, and filtered. After evaporation under vacuum, a colorless oil formed which was triturated with cyclohexane to form a white solid which was recrystallized from cyclohexane (yield: 83%); m.p. 148-150° C (uncorrected). NMR (in CDCl₃) δ 7.74-6.82 (multiplet, 9H, ArH); 4.68 (singlet, 2H, CH₂ in imidazolone ring); 3.61 (singlet, 1H, OH exchangeable with D₂O); 2.99-1.53 (multiplet, 18H). IR (KBr) 1680 cm⁻¹. Elemental analysis for C₂₄H₂₈N₃O₂F - Theoretical: C: 70.05, H: 7.35, N: 10.21. Found: C: 69.92, H: 7.39, N: 10.16.

ACKNOWLEDGEMENTS

This work was supported in part by the USPHS grant numbers MH-42821 and NS-15080.

REFERENCES

- 1. Forsman A. and Larsson M. Curr. Ther. Res. 24: 567 (1978).
- 2. Korpi E. R. and Wyatt R. J. Psychopharmacol. 83: 34 (1984).
- Korpi E. R., Costakos D. T. and Wyatt R. J. Biochem. Pharmacol. <u>34</u>: 2923 (1985).
- Chakraborty B. S., Hubbard J. W., Hawes E. M., McKay G., Cooper J. K., Gurnsey T., Korchinski E. D., Midha K. K. - Eur. J. Clin. Pharmacol. <u>37</u>: 45 (1989).
- Ereshefsky L., Davis C. M., Harrington C. A., Jann M. W., Browning J. L., Saklad S. R.and Burch N. R. - J. Clin. Psychopharmacol. <u>4</u>: 138 (1984)
- 6. Altamura A. C., Mauri M. C., Cavallaro R. and Gorni A. Lancet 814 (1987).
- 7. Miller A. K., Minkin E., Wong D. F., Young D. Psychopharmacol. submitted 1990.
- Shiue C.-Y., Fowler J. S., Wolf A. P., Shea C., and Adam M. J. J. Labelled Comp. Radiopharm. <u>26</u>: 386 (1989).
- Shiue C.-Y., Fowler J. S., Wolf A. P., McPherson D. W., Arnett C. D., and Zecca L. - J. Nucl. Med. <u>27</u>: 226 (1986).
- Kilbourn M. R. Fluorine-18 Labeling of Radiopharmaceuticals, Nuclear Science Series - Nuclear Medicine; National Academy Press; Washington, DC, p 17 (1990).
- 11. Janssen P. A. J., U.S. Patent No. 3155670, 1964.

- Burns H. D., Dannals R. F., Långström B., Ravert H. T., Zemyan S. E., Duelfer T., Wong D. F., Frost J. J., Kuhar M. J., Wagner H. N. Jr. - J. Nucl. Med. <u>25</u>, 303 (1985).
- Dannals R. F., Ravert H. T., Wilson A. A., Wagner H. N. Jr. Int. J. Radiat. Appl. Instrum. Part A, Appl. Radiat. Isot. <u>37</u>: 433 (1986).